Engineering of Allogeneic Regulatory T Cells Expressing a Chimeric Antigen Receptor (Allo-CAR-Tregs) Using Zinc Finger Nuclease/AAV6-Mediated Editing

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Introduction

- Conversion of polyclonal regulatory T cells (Tregs) into antigen-specific Tregs by the introduction of a chimeric antigen receptor (CAR) has gained increasing attention as a potential treatment option for autoimmune diseases.
- Designed to recognize a clinically relevant antigen, CAR-Tregs can be specifically activated through the CAR upon binding to a specific antigen, proliferate, and acquire their full immunomodulatory capacities. Therefore, these genetically modified CAR-Tregs are potentially able to prevent or dampen autoimmune reactions in patients.
- Allogeneic CAR-Tregs were engineered using a donor DNA consisting of a CAR-expressing adenoassociated virus type 6 (CAR-AAV6) and zinc finger nuclease (ZFN) editing technology to insert the CAR into the beta-2 microglobulin (B2M) locus, leading to the knockout of cell-surface expression of the major histocompatibility complex I (MHC-I). MHC-I plays a major role in allogeneic rejection, and by deleting it, we aim to limit the elimination of allogeneic CAR-Tregs by the patient's immune system.
- Here we generated CAR-Tregs using ZFN/AAV6 technology to screen various expression cassettes for optimal CAR expression and functionality. The tested CAR-Tregs were designed to recognize the human interleukin-23 (IL-23) receptor (IL23R), a key player in the pathogenesis of inflammatory bowel disease (IBD), and against the human myelin oligodendrocyte glycoprotein (MOG), an autoimmune target in multiple sclerosis (MS).

Tregs: a new class of cell-based therapeutics



- Tregs maintain immune homeostasis at various tissues.
- The suppressive function of Tregs inhibits the mounting of inflammatory responses, i.e., Tregs confer tolerance.
- Tregs can be used as a cell-based therapy across various applications where induction of immune tolerance can restore homeostasis and counter the disease state (e.g., prevention of transplant rejection, treatment of a multitude of autoimmune diseases).



- Allogeneic CAR-Treg cell product confers an advantage when autologous Tregs cannot be isolated from the patient.
- Universal off-the-shelf allogeneic CAR-Treg product will significantly cut costs and time.
- ZFN technology allows the development of allogeneic CAR-Tregs.

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Methods



Schematic showing the experimental design. Tregs are isolated from a healthy donor, followed by ex vivo cell engineering to knock out the expression of MHC-I and to introduce CAR into the B2M locus. InDels, insertions/deletions; TI, targeted integration.

AAV6 CAR expression vectors with different promoters

			→	
B2M-L	pА	PGK	IL23R/MOG-CAR	pA B2M-R
B2M-L	pА	PGK100	IL23R/MOG-CAR	pA B2M-R
				· · · ·
B2M-L	pА	EFS	IL23R/MOG-CAR	pA B2M-R
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Tregs were electroporated with B2M ZFN mRNA followed by AAV6 transduction (1) using the indicated vectors expressing either IL23R-CAR or MOG-CAR under the control of various promoters (human phosphoglycerate kinase [PGK], short PGK [PGK100], human elongation factor 1 alpha [EF1a], short EF1a [EFS], and short human FOXP3derived promoters [hFXP3.1 and hFXP3.2]).

Results

B2M ZFNs efficiently knock out MHC-I expression in Tregs



Knockout of MHC-I expression in Tregs using ZFN directed against the B2M gene locus. (A) Schematic representation of the MHC-I molecule composed of alpha 1, 2, and 3, and B2M proteins. (B) Viability of edited and non-edited Treg cells with B2M ZFN was assessed by fluorescent activated cell sorter analysis with propidium iodide staining at day 4 post electroporation. (C) Knockout of MHC-I expression after editing using B2M ZFN in human Tregs. Results are presented as bars and whiskers (n=8).

Image created with biorender.com

B2M-L	pА	hFXP3.1	IL23R/MOG-CAR	pA B2M-R
B2M-L	pА	hFXP3.2	IL23R/MOG-CAR	pA B2M-R
B2M-L	pА	EF1a	IL23R/MOG-CAR	pA B2M-R
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Screening of different promoters to modulate IL23R-CAR and MOG-CAR expression in Tregs. (A, D) Targeted integration (TI) of IL23R-CAR and MOG-CAR sequences into the B2M locus was quantified by next-generation sequencing (MiSeq). (B, E) The impact of the different promoters on CAR-Treg generation efficiency was assessed by measuring the percentage of CAR+/MHC-I(-) cells using flow cytometry. (C, F) IL23R-CAR and MOG-CAR expression levels on the Treg cell surface were determined by measuring the mean fluorescence intensity (MFI) of the CARs. Data are 3 independent experiments (n=9 and 8 Treg donors, respectively).

Highly efficient CAR-mediated activation of CAR-Tregs generated using **ZFN/AAV6 technology**



The CAR-mediated activation assay monitors the expression of the CD69 activation marker at the cell surface of CAR-Tregs. Activation of CAR-Tregs was measured 24 hours after incubation either with anti-CD3/CD28 beads to stimulate the T-cell receptor (positive control) or with IL23R-coated beads to stimulate the CAR (n=4). The ranking from optimal to minimal CAR-mediated activation is as follows: EF1a=PGK=EFS>PGK100=hFXP3.1>hFXP3.2.

Conclusions

- regardless of the expression cassette used.

- with AAV6-CAR donor DNA.

References

1. Wang J. *et al.* (2015) Nat. Biotechnol. 33(12):1256-1263.

Author Disclosures

All authors are or were employees of Sangamo Therapeutics

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Tregs engineered with ZFN/AAV6 technology at B2M locus efficiently express IL23R-CAR and MOG-CAR





 Our results show high MHC-I knockout efficiency ranging between 70%-90% with no impact on Treg viability. • The targeted integration efficiency was similar overall (40% to 50%) between all the AAV6-CAR donor DNA

Treas engineered with ZFN/AAV6 technology at the B2M locus efficiently express IL23R-CAR and MOG-CAR. • CAR-mediated activation was highly efficient, especially for CAR-AAV6 harboring the EF1a or PGK promoters. • Overall, our results provide a robust strategy for genome engineering in Tregs using ZFN in combination